THE 2-PHENYLSULFONYLETHYLCARBONYL (PSEC) GROUP FOR THE PROTECTION OF THE HYDROXYL FUNCTION

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Abstract: The PSEC group may be used to protect the hydroxyl function in conjunction with a variety of acid and base labile protecting groups; the PSEC group may be removed by treatment of triethylamine (15 equiv.) in dry pyridine solution within 20 h at 20°C while other base labile protecting groups remained intact.

The protection of hydroxyl function constitute an important step in designing a strategy for a multistep chemical synthesis of oligosacharides and oligonucleotides. Several acid, 1 base labile protecting groups  $^{1}$  along with the ones which are removable under virtually neutral conditions<sup>2,3</sup> have been proposed for this purpose. There are other categories of hydroxyl protecting groups which are removable either by mild lewis acids, 4,5 by fluoride ions 5,6 or by other means 7,8. We now report that the 2-phenylsulfonylethylcarbonyl (PSEC) group, as shown in  $(\underline{2})$ , a new hydroxyl protecting group which is removable quantitatively within 20 h at 20°C by the action of Et<sub>3</sub>N, a non-nucleophilic base, (15 equiv.) in dry pyridine solution (15 ml/mmol). Alternatively, the PSEC group from (2) may be removed by mild alkaline hydrolytic conditions. Thus it is completely removable with M-NH $_3$  in dioxan-water (1:1 v/v) within 7 min. at  $20^{\circ}\text{C}$  or with 0.04 M  $\text{K}_2\text{CO}_3$  in aqueous dioxan (1:1 v/v) at  $20^{\circ}\text{C}$  within a minute. The PSEC-group has the expected acid stability for over 10 days at 20°C in pH 2 (80% aq. acetic acid). The PSEC-derivative, as in (2), can be conveniently prepared by treating a pyridine solution (10 m1/mmol) of the substrate at 20°C with 2-phenylsulfonylethylchloroformate<sup>9</sup> (1) (1.3 equiv.) for 30 min. followed by a standard work-up. The Table illustrates the substrates (3) to (9), containing different standard protecting groups, whose PSEC-derivatives have been prepared.

The Table also records the % isolated yields: (i) of formation of the PSEC-derivatives  $^{18}$  of the substrates (3) to (9); (ii) of the recovery of the substrates  $^{18}$  after the removal of the PSECgroup in presence of other protecting groups; (iii) and of the recovery yields of the hydroxy block containing the PSEC-group after the deprotection of the corresponding acid or base labile protecting group from the PSEC-derivatives  $^{18}$  of (3) to (7). Thus, the examples in the Table clearly establish the synthetic applicability of the PSEC-group in conjunction with acid labile groups, like 9-phenylxanthen-9-yl (Pixyl)10 and tetrahydropyranyl1, and base labile groups like  $benzoy1^1$ , 2,2-dibromomethylbenzoy1<sup>2</sup>, 2-trimethylsilylethylcarbonyl-(TMSEC)<sup>5</sup> and levulinyl-<sup>3</sup>. The selective removal of the PSEC-group from the PSEC-derivative of the substrate (4) is particularly interesting in view of the fact that the DBMB-group  $^2$  has an almost identical lability to that of an acetyl group in an alkaline hydrolytic condition  $^{11}$  (t<sub>1/2</sub> Ca. 90 min. at 20 °C 0.15M aq.  $K_2CO_3$  in dioxan solution, 1.6:1.0 v/v). It is also noteworthy that the selective removal of the DBMB-group by a virtually neutral condition using AgClO4 (16 equiv.),2,4,6collodine (9 equiv.) in 98% aq. acetone for 1 h. followed by the treatment with morpholine (5 equiv.), the removal of the TMSEC-group  ${}^{5,19}(t_{1/2}$  ca. 7 h., dioxan-aq. NH $_3$  (d0.9), 1:1 v/v by the action of ZnBr<sub>2</sub> (10 equiv.) in CH<sub>3</sub>NO<sub>2</sub> solution at 20°C and the removal of levulinyl

TABLE: PREPARATION OF THE PSEC-DERIVATIVES AND THE RECOVERY YIELDS.

SUBSTRATES	The formation of the PSEC-deriva- tives.	removal of the PSEC-group in presence of other groups a.	The selective removal of other groups in presence of the PSEC-group.
	% Yields	% Yields	% yields
$(\underline{3}) \qquad \qquad \bigcap_{O} Ph \\ O \\ O \\ OH$	86.6	85.8	95.6
$(\underline{4}) \qquad \qquad \bigcirc CHBr_2 \\ O \qquad OH \qquad CBz$	99.0	98.2	73.6
$(\underline{5}) \qquad \qquad \bigcap_{O} \bigcap_{T} \bigcap_{T} \bigcap_{O} \bigcap_{T} \bigcap_{T} \bigcap_{O} \bigcap_{T} \bigcap_{T}$	80.6	86.1	92.0
$(\underline{6}) \qquad \qquad \downarrow 0 \qquad \uparrow \qquad \qquad \downarrow 0$	88.0	87.3	96.0
$(7) \qquad \qquad \begin{array}{c} \text{O} \qquad \qquad \text{SiMe}_3 \\ \text{O} \qquad \qquad \\ \text{O} \qquad \qquad$	80.0	99.0	96.7
$(8) \qquad \qquad \downarrow 0 \qquad \downarrow T \qquad \qquad \downarrow 0$	73.6 b	84.7 c	v
он ( <u>9</u> ) CHOLESTEROL	74.0	96.5 d	

 $<sup>^{</sup>a}$   $_{15}$  equiv. of Et $_{3}$ N in dry pyridine(15 ml/mmol) was used for deprotection of the PSEC-group (20h)  $_{5}$ '-0-PSEC derivative was selectively obtained.

The deprotection took 70h under the usual condition.

 $<sup>^{\</sup>rm C}$  The 5'-0-PSEC derivative of (8) was first benzoylated at 3'-position and then the PSEC-group  $^{\rm C}$  was removed to yield 3'-0-benzoyl thymidine.

group  $^3$  (0.5 M hydrazine hydrate in pyridine-acetic acid, 3:1 v/v at 20°C) from the PSEC-derivatives of  $(\underline{4})$ ,  $(\underline{6})$  and  $(\underline{7})$  respectively should broaden the potential use of the PSEC-group in the synthesis of the natural products.

Finally, the application of the PSEC-group in oligodeoxyribonucleotide synthesis was investigated using a reported strategy. 12 Thus, the dimer block (10) was synthesized in 71.4% yield (powder) by reacting the 5'-0-PSEC thymidine (9) with a slight excess of o-chlorophenylphosphorobis-(1,2,4-triazolide) in CH3CN-pyridine solution at 20°C for 30 min in an usual fashion followed by an addition thymidine (2 equiv.) and 1-methylimidazole (16 equiv.). $^{13}$ The reaction mixture was stirred for 45 min followed by a standard work-up and purification by column chromatography gave the pure (10). The 3'-hydroxyl function of the (10) was then blocked with the pixyl group $^{10}$  to obtain (11) in a quantitative yield. The 5'-hydroxy dimer block (12) was then prepared in 94% yield by the removal of the PSEC-group by the treatment of EtaN (15 equiv.) in dry pyridine solution (15 ml/mmol) for 20 h. at 20°C. The 5'-protected dimer block (13) was conveniently prepared following a literature procedure. 14 A slight excess of the (13) was then reacted with the (12) under a usual condition in presence of an excess of 1-mesitylenesulfonyl-3-nitro-1,2,4-triazole<sup>12,15</sup> to obtain the fully protected tetramer (14) in 94.4% yield. The tetramer (14) gave a pure 5'-hydroxy block (16) in 81% yield (powder), after the removal of the PSEC-group using the above Et3N in pyridine reaction condition and a pure 5'-protected phosphodiester component (15) following a literature procedure  $^{12,16}$ . The (16)was then coupled to the (15) in a standard way to obtain the fully protected octathymidylic acid (17) in 84% yield (powder). It should be added here that the 0.04M aq. K2CO3 in dioxan (1:1 v/v, 20°C, 6 min.  $T_2$  &  $T_4$  and 8 min for  $T_8$ ) removed the PSEC-group from the fully protected dimer (11), tetramer (14) and the octamer (17) to their 5'-hydroxy components in 82.4, 64.3 and 47.7% yields as opposed to Et<sub>3</sub>N (15 equiv.) in dry pyridine (20 h. at 20°C) condition which gave the respective 5'-hydroxy blocks in 94, 81 and 80.6% yields. The poorer yield of 5'-hydroxy components in the former conditions was clearly due to the removal of o-chlorophenyl group from

(10) R=H, R<sup>1</sup>=PSEC

(12) R=Px, R<sup>1</sup>=H

$$(13) R = \underbrace{0 - C1Ph0}_{P} = \underbrace{0 - Et_3NH}^{+}$$

$$R^{1} = PSEC$$

 $(\underline{14})$  n=2,  $R_1^1$ =PSEC, R=Px

(15) n=2, 
$$R^1$$
=PSEC and  $R=$ 
0-C1Ph0
 $P = 0$ 
 $Et_3$ 

 $(\underline{16})$  n=2,  $R^{1}$ =H and R=Px

(17) n=6,  $R^1$ =PSEC and R=Px

from the internucleotide junctions. The final product ( $\underline{17}$ ) was deprotected in the following order: (1) replacement of the benzoyl group  $^{16}$  at the 5'-end after the removal of the PSEC-group; (ii) treatment with  $\underline{\text{syn}}$ -4-nitrobenzaldoximate  $ions^{15}$ ; (iii) aq. NH<sub>3</sub> (d0.9) for 24h. at 20°C; (iv) removal of the volatile matters and the treatment of the residue with 80% aq.acetic acid for 15 min. at 20°C. The oligonucleotide was then extracted into the aqueous phase by partitioning the reaction mixture with CH<sub>2</sub>Cl<sub>2</sub> (10x5ml). The aqueous phase was then concentrated and examined by HPLC<sup>16</sup> (Permaphase AAX at 50°C; linear gradient: 0.01M KH<sub>2</sub>PO<sub>4</sub>, 0.0M KCl to 0.05M KH<sub>2</sub>PO<sub>4</sub> and 0.7M KCl; pH 4.45) and the main peak in the elution profile contained more than 95% of the  $A_{260}$  0.D. units. An aliquot of this material was  $^{32}$ P-labelled with  $^{32}$ P- $^{3}$ -ATP and kinase and electrophoresed on a 20% polyacrylamide gel<sup>17</sup>. An autoradiography revealed the presence of a single component of the expected mobility. The oligonucleotide was fully digested with Crotalus adamanteus snake venom phosphodiesterase confirming the presence of 3' $\Rightarrow$ 5' linkages in the octathymidylic acid. The  $^{32}$ P-labelled octamer was also partially digested by the same enzyme and then it was electrophoretically shown to contain eight fragments of differently charged species as expected from octathymidylic acid<sup>17</sup>.

Thus, it is clearly demonstrated that the PSEC group can indeed be successfully used as a hydroxyl protecting group. The obvious advantage with the PSEC-group seem to lie in the fact that one could conveniently introduce either electron withdrawing or lipophilic substituent, or a sutiable combination of the both, in the benzene ring in order to make the substituted 2-Phenylsulfonylethyl linkage more labile to a non-nucleophilic base to suit other needs.

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## References and notes:

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1. C.B. Reese, Protection of alcoholic hydroxyl groups and Glycols systems in "Protective Groups
in Organic Chemistry", ed. J.F.W. McOmie, Plenum, 1973.
2. J.B. Chattopadhyaya, C.B. Reese and A.R. Todd, J.C.S.Chem.Comm. 987 (1979).
3. J.H. van Boom and P.M.J. Burgers. Tetrahedron Lett. 4875 (1976).
4. E.J. Corey, J.-L. Gras and P. Ulrich Tetrahedron Lett. 809 (1976).
   V. Kohli, H. Blöcker and H. Köster. Tetrahedron Lett. 2683 (1980).
   M.D. Metteucci and M.H. Caruthers. Tetrahedron Lett. 3243 (1980).
5. C. Gioeli, N. Balgobin, S. Josephson and J.B. Chattopadhyaya. Tetrahedron Lett. 969 (1981).
6. E.J. Corey and A. Venkateswazlu. J.Am. Chem. Soc. 94, 6190 (1972).
   B.H. Lipshutz and J.J.Pergram. Tetrahedron Lett. 3343 (1980).
   W.T. Markiewicz. J. Chem. Res. (S) 24 (1979).
7. P.M. Pojer and S.J. Angyal. Tetrahedron Lett. 3067 (1976).
   T.-L. Ho and T.W. Hall. Synthetic Comm. 5(5), 367 (1976).
8. S.S.Jones, C.B. Reese and S. Sibanda. Tetrahedron Lett. 1933 (1981).
9. The reagent (1) has been conveniently prepared by reacting the 2-phenylsulfonylethanol (H.S.
   Schultz et.al. J.Org.Chem. 28, 1141 (1963) with an excess of phosgene in toluene solution
   according to the procedure described by V.P. Kozynkov et al. Zhur Obshcheie Khim, 38, 1179
   (1968). The chloroformate (1) spontaneously decomposed on attempts to purify by distillation
   in vacuo. Therefore, the crude (1) was used as a reagent which was sufficiently pure
   derivatization. H-NMR(CDC13):87.9-7.4 (5 H, aromatic protons), 4.62 (t, 2H, -CH2-OCCC1
   3.52 (\underline{t}, 2H, PhSO<sub>2</sub>-CH<sub>2</sub>-); I.R. (film): \hat{y}-0-COC1 1773 cm<sup>-1</sup>; 1305, 1288, 1130 cm<sup>1</sup>. ),
10.J.B.Chattopadhyaya and C.B. Reese. J.C.S.Chem.Comm. 639 (1978). 11.J.B.Chattopadhyaya and C.B. Reese. Unpublished result.
12.J.B.Chattopadhyaya and C.B. Reese. Nucleic Acid Res. 8, 2039 (1980).
13.S. Josephson, N. Balgobin and J.B. Chattopadhyaya. Proc. 4th Round table,
   Nucleosides, Nucleotides & their Biological applications, Antwerp, Belgium, 1981, p.7.
14.J.B.Chattopadhyaya and C.B. Reese. <u>Tetrahedron Lett.</u> 5059 (1979). 15.C.B.Reese, R.C. Titmus and L. Yau, <u>Tetrahedron Lett.</u> 2727 (1978).
16.N.Balgobin, S.Josephson and J.B. Chattopadhyaya. Acta Chem. Scand. Ser. B. (1981) in press.
17.E.Jay, R.Bambara, R. Padmanabhan and R. Wu. Nucleic acid Res. 1, 331 (1974).
18.The products have been characterized by <sup>1</sup>H-NMR, I.R., mass spectrometry and microanalyses.
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19.F ions promoted deprotection of the TMSEC-group led to an extensive cleavage of the PSEC-

group (see ref.5 for deprotection condition).